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### Research Article

### Biochemistry

## Reactive Oxygen Species Scavenging Activity of *Furcraea foetida* Leaf – An *In Vitro* Study

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### ABSTRACT

Antioxidant activity of *Furcraea foetida* leaf extract were carried out for proving its utility in free radical mediated diseases including diabetic, cardiovascular, cancer etc. The ethanolic extract was screened for *in vitro* antioxidant activity by oxygen radical scavenging such as DPPH, total antioxidant assay, superoxide, metal chelation and iron reducing power activity at different concentrations. Throughout the studies leaf extract showed marked antioxidant activity. The antioxidant activity of the leaf extract may be due to the phytochemicals present in it. The antioxidant activity was found to be concentration dependent and may be attributed to the presence of bioflavonoids content in *Furcraea foetida* leaf extract (FFLE). Overall, the plant extract is a source of natural antioxidants which might be helpful in preventing the progress of various oxidative stress mediated diseases including aging.

**Keywords:** Antioxidant activity, *Furcraea foetida* leaf, Radical scavenging, Reactive oxygen species,

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### INTRODUCTION

In living organisms, various reactive oxygen and nitrogen species (ROS/RNS) e.g., superoxide anions ( $O_2^{\cdot-}$ ), hydroxyl radicals ( $\cdot OH$ ), nitric oxide radicals ( $NO\cdot$ ) and non-radical compounds, can be formed by different mechanisms. It is unavoidable one because of they are continuously produced by the body's normal use of oxygen. Such species are considered to be important causative factors in the development of diseases such as diabetes, stroke, arteriosclerosis, cancer, and cardiovascular diseases and the aging process (Velavan, 2011; Alma *et al*, 2003). This effect was significantly reversed by prior administration of antioxidant providing a close relationship between free radical scavenging activity (FRSA) and the involvement of endocrinological responses (Wiseman and Halliwell, 1996).

The recent abundant evidence suggesting the involvement of oxidative stress in the pathogenesis of various disorders and diseases has attracted much attention of the scientists and general public to the role of natural antioxidants in the maintenance of human health and prevention and treatment of diseases (Niki, 2010). Plant and its products are rich sources of a phytochemicals and have been found to possess a variety of biological activities including antioxidant potential (Velavan *et al.*, 2007). The majority of the active antioxidant constituents are flavonoids, isoflavones, flavones, anthocyanins, coumarins, lignans, catechins, and isocatechins. In addition to the above compounds found in natural foods, vitamins C and E, beta-carotene, and tocopherol are known to possess antioxidant potential (Prior, 2003). With this background and abundant source of unique active components harbored in plants. The chosen medicinal plant namely as *Furcraea foetida* (Tamil: Annaikatalai) leaf belongs to the Asparagaceae family. Therefore, the present study were to investigate the free radical scavenging activity of *Furcraea foetida* leaf through the free radical scavenging such as DPPH scavenging, superoxide anion radical scavenging, metal chelation, reducing power activity and total antioxidant assay.

## MATERIALS AND METHODS

### Chemicals

Nitro blue tetrazolium (NBT), ethylene diamine tetra acetic acid (EDTA), sodium nitroprusside (SNP), trichloro acetic acid (TCA), thio barbituric acid (TBA), potassium hexa cyano ferrate [ $K_3Fe(CN)_6$ ], and L-ascorbic acid were purchased from Sisco Research Laboratories Pvt. Ltd., India. All other chemicals and solvents used were of analytical grade available commercially.

### Plant materials:

The fully mature *Furcraea foetida* leaves were collected in April 2014 from Tamil University, Thanjavur District, Tamil Nadu, India from a single herb. The leaves were identified and authenticated by Botanist, Dr. S John Britto, Department of Botany, St. Josephs College, Tiruchirappalli, Tamil nadu, India. A Voucher specimen has been deposited at the Rapinat Herbarium, St. Joseph,s College, Tiruchirappalli, Tamil nadu, India.

### Plant sample extraction

20gm powdered plant material is soaked in 50ml of Absolute alcohol overnight and then filtered through Whatmann filter paper No.41 along with 2gm sodium sulfate to remove the sediments and traces of

water in the filtrate. Before filtering, the filter paper along with sodium sulphate is wetted with absolute alcohol. The filtrate is then concentrated by bubbling nitrogen gas into the solution and reduce the volume to 1ml. The extract contains both polar and non-polar phytochemicals. Doses such as 20, 40, 60 and 80µg/ml were chosen for *in vitro* antioxidant activity.

### *In vitro* antioxidant activity

#### DPPH ASSAY

The scavenging ability of the natural antioxidants of the plant extract towards the stable free radical DPPH was measured by the method of Shimada *et al.* (1992). Briefly, a 2 ml aliquot of DPPH methanol solution (25µg/ml) was added to 0.5 ml sample solution at different concentrations. The mixture was shaken vigorously and allowed to stand at room temperature in the dark for 30 min. Then the absorbance was measured at 517nm in a spectrophotometer. L-Ascorbic acid was used as the standard.

$$\text{Radical scavenging activity (\%)} = 100 - \frac{A_C - A_S}{A_C} \times 100$$

Where  $A_C$  = control is the absorbance of the control and  $A_S$  = sample is the absorbance of reaction mixture (in the presence of sample). All tests were run in triplicates (n = 3), and the average values were calculated.

#### Determination of total antioxidant capacity

The antioxidant activity of the extracts was evaluated by the phosphomolybdenum method according to the procedure of Prieto *et al.* (1999). The assay is based on the reduction of Mo(VI)–Mo(V) by the extract and subsequent formation of a green phosphate/Mo(V) complex at acid pH. 0.3 ml extract was combined with 3ml of reagent solution (0.6M sulfuric acid, 28mM sodium phosphate and 4mM ammonium molybdate). The tubes containing the reaction solution were incubated at 95°C for 90 min. Then the absorbance of the solution was measured at 695 nm using a spectrophotometer against blank after cooling to room temperature. Methanol (0.3 ml) in the place of extract is used as the blank. The antioxidant activity is expressed as the number of equivalents of ascorbic acid.

#### Superoxide anion scavenging activity assay

The scavenging activity of the *Furcraea foetida* towards superoxide anion radicals was measured by the method of Liu *et al.* (1997). Superoxide anions were generated in a non-enzymatic phenazine methosulfate-nicotinamide adenine

dinucleotide (PMS-NADH) system through the reaction of PMS, NADH, and oxygen. It was assayed by the reduction of nitroblue tetrazolium (NBT). In these experiments the superoxide anion was generated in 3 ml of Tris-HCl buffer (100mM, pH 7.4) containing 0.75 ml of NBT (300  $\mu$ M) solution, 0.75 ml of NADH (936  $\mu$ M) solution and 0.3 ml of different concentrations of the extract. The reaction was initiated by adding 0.75 ml of PMS (120  $\mu$ M) to the mixture. After 5 min of incubation at room temperature, the absorbance at 560 nm was measured in spectrophotometer. The superoxide anion scavenging activity was calculated according to the following equation:

$$\% \text{ Inhibition} = [(A_0 - A_1) / A_0 \times 100],$$

Where  $A_0$  was the absorbance of the control (blank, without extract) and  $A_1$  was the absorbance in the presence of the extract.

### Fe<sup>2+</sup> chelating activity assay

The chelating activity of the extracts for ferrous ions Fe<sup>2+</sup> was measured according to the method of Dinis *et al.* (1994). To 0.5 ml of extract, 1.6 ml of deionized water and 0.05 ml of FeCl<sub>2</sub> (2 mM) was added. After 30 s, 0.1 ml ferrozine (5 mM) was added. Ferrozine reacted with the divalent iron to form stable magenta complex species that were very soluble in water. After 10 min at room temperature, the absorbance of the Fe<sup>2+</sup>-Ferrozine complex was measured at 562 nm. The chelating activity of the extract for Fe<sup>2+</sup> was calculated as

$$\text{Chelating rate (\%)} = (A_0 - A_1) / A_0 \times 100$$

Where  $A_0$  was the absorbance of the control (blank, without extract) and  $A_1$  was the absorbance in the presence of the extract.

### Reducing power assay

The Fe<sup>3+</sup> reducing power of the extract was determined by the method of Dinis (1994) with slight modifications. The extract (0.75 ml) at various concentrations was mixed with 0.75 ml of phosphate buffer (0.2 M, pH 6.6) and 0.75 ml of potassium hexacyanoferrate [K<sub>3</sub>Fe(CN)<sub>6</sub>] (1%, w/v), followed by incubating at 50°C in a water bath for 20 min. The reaction was stopped by adding 0.75 ml of trichloroacetic acid (TCA) solution (10%) and then centrifuged at 3000 r/min for 10 min. 1.5 ml of the supernatant was mixed with 1.5 ml of distilled water and 0.1 ml of ferric chloride (FeCl<sub>3</sub>) solution (0.1%, w/v) for 10 min. The absorbance at 700 nm was measured as the reducing power. Higher absorbance of the reaction mixture indicated greater reducing power.

**Statistical analysis:** Tests were carried out in triplicate for 3–5 separate experiments. The amount of extract

needed to inhibit free radicals concentration by 50%, IC<sub>50</sub>, was graphically estimated using a nonlinear regression algorithm.

## RESULTS AND DISCUSSION

The search for new molecules, nowadays, has taken a slightly different route where the science of ethnobotany and ethnopharmacognosy are being used as guide to lead the chemist towards different sources and classes of compounds (Gurib-Fakim, 2006). Plant derived natural products hold great promise for discovery and development of new pharmaceuticals (McChesney *et al.*, 2007). Our earlier reports indicates tha methanolic extract of *Furcraea foetida* leaves extract contains flavonoids, saponin, terpenoids, steroids, polyphenols, saponin and triterpenoids which are an important in disease prevention and health preservation.

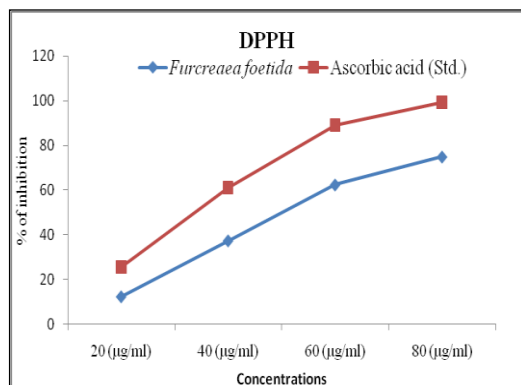
### DPPH Assay

Recently, the use of the DPPH<sup>•</sup> reaction has been widely diffused among food technologists and researchers, for the evaluation of free radical scavenging activity on extracts from plant, food material or on single compounds. In the DPPH assay, the antioxidant was able to reduce the stable radical DPPH to the yellow colored 1, 1-diphenyl-1, 2-picryl hydrazine. The molecule of 2, 2-diphenyl-1-picryl hydrazine is characterised as a stable free radical by virtue of the delocalisation of the spare electron over the molecule as a whole. The proton transfer reaction of the DPPH<sup>•</sup> free radical by a scavenger causes a decrease in absorbance at 517 nm, which can be followed by a common spectrophotometer set in the visible region. The effect of antioxidants on DPPH<sup>•</sup> is thought to be due to their hydrogen donating ability (Sindhu and Abraham, 2006). DPPH radical scavenging activity of plant extract of FFLE and standard as ascorbic acid are presented in Table 1 and Fig 1. The DPPH radical was widely used to evaluate the free-radical scavenging capacity of antioxidants (Nuutila *et al.*, 2003). The half inhibition concentration (IC<sub>50</sub>) of ascorbic acid and plant extract were 47.55 $\mu$ g ml<sup>-1</sup> and 52.96  $\mu$ g ml<sup>-1</sup> respectively. The plant extract exhibited a significant dose dependent inhibition of DPPH activity. The potential of L-ascorbic acid to scavenge DPPH radical is directly proportional to the concentration. The DPPH assay activity is near to standard as ascorbic acid.

**Table 1- % of DPPH Radical scavenging activity of *Furcraea foetida* extract at different concentrations**

Parameters	20 (µg/ml)	40 (µg/ml)	60 (µg/ml)	80 (µg/ml)	IC <sub>50</sub>
<i>Furcraea foetida</i> extract	12.5 ± 0.88	37.5 ± 2.65	62.5 ± 4.38	75.00 ± 5.25	52.96
Standard (Ascorbic acid)	25.6±2.04	61.26±4.90	88.98±7.11	99.34±7.94	47.55

Values were expressed as Mean ± SD for triplicates

**Fig 1- % of DPPH Radical scavenging activity of *Furcraea foetida* extract at different concentrations**

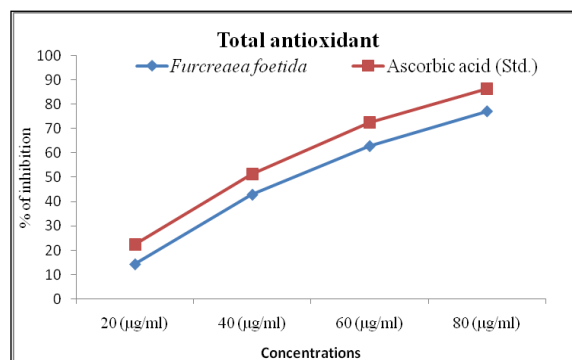
#### Total antioxidant activity

The phosphomolybdenum method was based on the reduction of Mo (VI) to Mo (V) by the antioxidant compound and the formation of a green phosphate/ Mo (V) complex with a maximal absorption at 695 nm. The assay is successfully used to quantify vitamin E in seeds and, being simple and independent of other antioxidant measurements commonly employed, it was decided to extend its application to plant extract (Prieto *et al.*, 1999). Moreover, it is a quantitative one, since the antioxidant activity is expressed as the number of equivalents of ascorbic acid. The yield of the ethanol extract of the plant extract and its total antioxidant capacity are given in Table 2 Fig 2. Total antioxidant capacity of FFLE is expressed as the number of equivalents of ascorbic acid. The study reveals that the antioxidant activity of the extract is in the increasing trend with the increasing concentration of the plant extract. The half inhibition concentration (IC<sub>50</sub>) of ascorbic acid and plant extract were 48.08 µg ml<sup>-1</sup> and 50.76 µg ml<sup>-1</sup> respectively.

**Table 2- % of Total antioxidant activity of *Furcraea foetida* extract at different concentrations**

Parameters	20 (µg/ml)	40 (µg/ml)	60 (µg/ml)	80 (µg/ml)	IC <sub>50</sub>
<i>Furcraea foetida</i> extract	14.2 ± 1.00	42.8 ± 3.00	62.8 ± 2.99	77.1 ± 4.00	50.76
Standard (Ascorbic acid)	22.35±1.80	51.23±4.09	72.54±5.80	86.35±6.91	48.08

Values were expressed as Mean ± SD for triplicates

**Fig 2- % of Total antioxidant activity of *Furcraea foetida* extract at different concentrations**

#### Superoxide anion radical scavenging activity

Superoxide is biologically important since it can be decomposed to form stronger oxidative species such as singlet oxygen and hydroxyl radicals, is very harmful to the cellular components in a biological system (Korycka-Dahl & Richardson, 1978). The superoxide anion radical scavenging activities of the extract from *Furcraea foetida* assayed by the PMS-NADH system were shown in Table 3 and Fig 3. The superoxide scavenging activity of *Furcraea foetida* was increased markedly with the increase of concentrations. The half inhibition concentration (IC<sub>50</sub>) of *Furcraea foetida* was 50.45 µg ml<sup>-1</sup> and ascorbic acid were 31.62 µg ml<sup>-1</sup> respectively. These results suggested that *Furcraea foetida* had notably superior superoxide radical scavenging effects.

#### The ferrous ion chelating activity

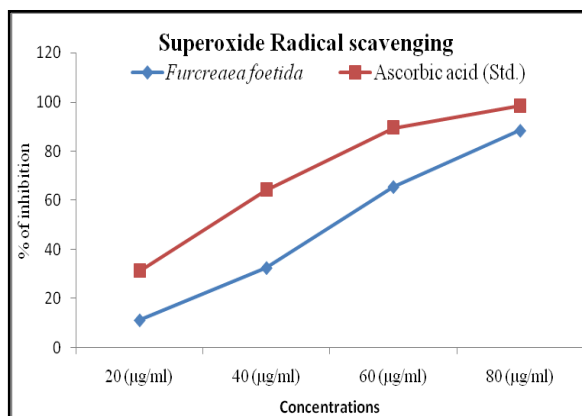
Ferrozine can make complexes with ferrous ions. In the presence of chelating agents, complex (red colored) formation is interrupted and as a result, the red color of the complex is decreased. Thus, the chelating effect of the coexisting chelator can be determined by measuring the rate of color reduction. The formation of

the ferrozine– Fe<sup>2+</sup> complex is interrupted in the presence of aqueous extract of *Furcraea foetida*, indicating that have chelating activity with an IC<sub>50</sub> of 54.87 µg ml<sup>-1</sup> and ascorbic acid was 30.96µg ml<sup>-1</sup> respectively (Table 4 and Fig 4). Ferrous iron can initiate lipid peroxidation by the Fenton reaction as well as accelerating peroxidation by decomposing lipid hydroperoxides into peroxy and alkoxy radicals (Halliwell, 1991; Fridovich, 1995). Metal chelating activity can contribute in reducing the concentration of the catalyzing transition metal in lipid peroxidation. Furthermore, chelating agents that form s bonds with a metal are effective as secondary antioxidants because they reduce the redox potential, and thereby stabilize the oxidized form of the metal ion (Gordon, 1990). Thus, *Furcraea foetida* demonstrate a marked capacity for iron binding, suggesting their ability as a peroxidation protector that relates to the iron binding capacity.

**Table 3- % of Superoxide Radical scavenging activity of *Furcraea foetida* extract at different concentrations**

Parameters	20 (µg/ml)	40 (µg/ml)	60 (µg/ml)	80 (µg/ml)	IC <sub>50</sub>
<i>Furcraea foetida</i> extract	11.24 ±0.78	32.54± 2.32	65.52 ± 4.67	88.43 ± 6.22	50.45
Standard (Ascorbic acid)	31.25 ±2.50	64.23 ± 5.13	89.54 ± 7.16	98.51 ± 7.88	31.62

Values were expressed as Mean ± SD for triplicates

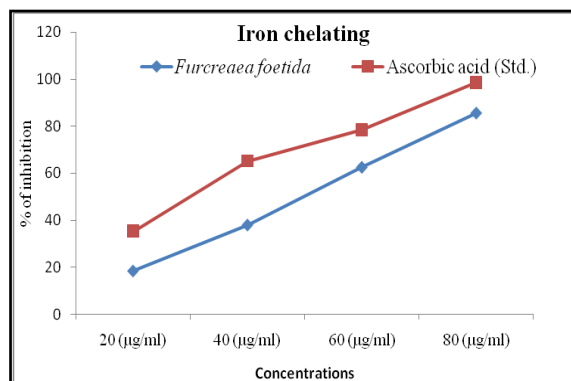


**Fig 3- % of Superoxide Radical scavenging activity of *Furcraea foetida* extract at different concentrations**

**Table 4- % of Iron chelating activity of *Furcraea foetida* extract at different concentrations**

Parameters	20 (µg/ml)	40 (µg/ml)	60 (µg/ml)	80 (µg/ml)	IC <sub>50</sub>
<i>Furcraea foetida</i> extract	18.5 ± 0.88	38.00 ± 3.5	62.5 ± 4.38	85.50 ± 4.35	49.02
Standard (Ascorbic acid)	35.23 ± 2.81	65.21 ± 5.28	78.51± 6.28	98.65 ± 7.89	30.96

Values were expressed as Mean ± SD for triplicates



**Fig 4- % of Iron chelating activity of *Furcraea foetida* extract at different concentrations**

**Reducing power activity**

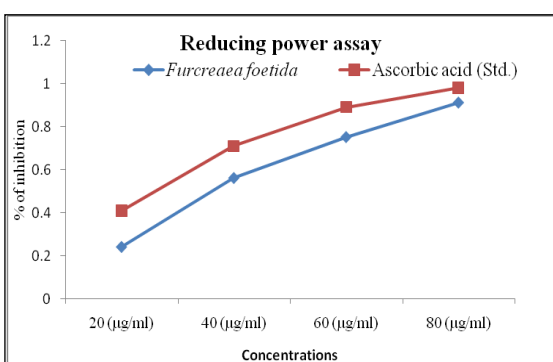
The measurements of the reducing ability, the Fe<sup>3+</sup>–Fe<sup>2+</sup> transformation was investigated in the presence of *Furcraea foetida*. The reducing capacity of a compound may serve as a significant indicator of its potential antioxidant activity. However, the activity of antioxidants has been assigned to various mechanisms such as prevention of chain initiation, binding of transition-metal ion catalysts, decomposition of peroxides, prevention of continued hydrogen abstraction, reductive capacity and radical scavenging (Diplock, 1997; Yildirim *et al*, 2000). Table 5 and Fig 3 depicts the reductive effect of *Furcraea foetida*. Similar to the antioxidant activity, the reducing power of *Furcraea foetida* increased with increasing dosage. All the doses showed significantly higher activities than the control exhibited greater reducing power, indicating that *Furcraea foetida* consist of hydrophilic polyphenolic compounds that cause the greater reducing power.



**Table 5- Reducing power assay of *Furcraea foetida* extract at different concentrations**

Parameters	20 (µg/ml)	40 (µg/ml)	60 (µg/ml)	80 (µg/ml)
<i>Furcraea foetida</i> extract	0.24±0.01	0.56±0.03	0.75±0.05	0.91±0.06
Standard (Ascorbic acid)	0.41± 0.03	0.71 ± 0.05	0.89± 0.07	0.98 ± 0.08

Values were expressed as Mean ± SD (Optical density) for triplicates

**Fig 5- Reducing power assay of *Furcraea foetida* extract at different concentrations**

## CONCLUSION

On the basis of the results of this study, it clearly indicates that *Furcraea foetida* leaf had powerful *in vitro* antioxidant capacity against various antioxidant systems as DPPH, nitric oxide, superoxide anion scavenging and metal chelator. From our results, the antioxidant activity of *Furcraea foetida* leaf was concentration dependent. The extracts could exhibit antioxidant properties approximately comparable to commercial synthetic antioxidants as ascorbic acid. From the above assays, the possible mechanism of antioxidant activity of *Furcraea foetida* leaf includes reductive ability, metal chelator, hydrogen donating ability and scavengers of superoxide and free radicals.

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## REFERENCES

- Alma MH, Mavi A, Yildirim A, Digrak M, Hirata T (2003). Screening chemical composition and antioxidant and antimicrobial activities of the essential oils from *Origanum syriacum* L. growing in Turkey. *Biol. Pharm. Bull.* 26: 1725–1729.
- Dinis TCP, Madeira VMC, Almeida LM (1994). Action of phenolic derivatives (acetoaminophen, salicylate, and 5-aminosalicylate) as inhibitors of membrane lipid peroxidation and peroxy radicals scavengers. *Achieves of Biochemistry and Biophysics*, 315: 161-169.
- Diplock AT (1997). Will the ‘good fairies’ please prove to us that vitamin E lessens human degenerative disease? *Free Radical Research*, 27: 511-532.
- Gordon MH (1990). The mechanism of the antioxidant action *in vitro*. In B. J. F. Hudson, *Food Antioxidants*, (pp. 1-18). London: Elsevier.
- Gurib-Fakim A. 2006. Traditions of yesterday and drugs of tomorrow. *Molecular Aspects of Medicine*. Special Edition. Elsevier Publications, UK.
- Halliwell B, (1991). *Free Radicals in Biology and Medicine*. (pp. 235-247). Oxford: Clarendon.
- Korycka-Dahl M, Richardson M (1978). Photogeneration of superoxide anion in serum of bovine milk and in model systems containing riboflavin and aminoacids. *Journal of Dairy Science*, 61: 400-407.
- Liu F, Ooi VEC, Chang ST (1997). Free radical scavenging activity of mushroom polysaccharide extracts. *Life Sci.* 60: 763-771.
- McChesney J., S. Venkataraman & J. Henri. 2007. Plant natural products: Back to the future or into extinction? *Phytochemistry* 68: 2015-2022.
- Niki E (2010). Assessment of Antioxidant Capacity *in vitro* and *in vivo*. *Review Article Free Radical Biology & Medicine* 49 503–515.
- Nuutila, A. M., Pimia, R. P., Aarni, M., & Caldenty, K. M. O. (2003). Comparison of antioxidant activities of onion and garlic extracts by inhibition of lipid peroxidation and radical scavenging activity. *Food Chemistry*, 81, 485–493.
- Okhawa H, Ohishi N, Yagi K (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical Biochemistry*, 95: 351-358.
- Oyaizu M (1986). Studies on products of browning reactions: antioxidant activities of products of browning reaction prepared from glucose amine. *Japanese Journal of Nutrition*, 44: 307-315.

- Prieto, P., Pineda, M., & Aguilar, M. (1999). Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: Specific application to the determination of vitamin E. *Analytical Biochemistry*, 269, 337–341.
- Prior RL (2003). Fruit and vegetables in the prevention of cellular oxidative damage. *American Journal of Clinical Nutrition*. 78 570S-578S.
- Shimada, K., Fujikawa, K., Yahara, K., & Nakamura, T. (1992). Antioxidative properties of xanthum on the autoxidation of soybean oil in cyclodextrin emulsion. *Journal of Agricultural and Food Chemistry*, 40, 945–948.
- Sindhu M, Abraham TE. (2006) In vitro antioxidant activity and scavenging effects of *Cinnamomum verum* leaf extract assayed by different methodologies. *Food and Chemical Toxicology* 44 198–206.
- Velavan S (2011). Free radicals in health and diseases- A Mini Review. *Pharmacologyonline Newsletter*. 1: 1062-1077.
- Velavan S, Nagulendran K, Mahesh R (2007). In vitro antioxidant activity of *Asparagus racemosus* leaf. *Pharmacog. Magaz*; 26-33.
- Wiseman H, Halliwell B (1996). Damage to DNA by reactive oxygen and nitrogen species: role of inflammatory disease and progression to cancer. *Biochem. J.* 313: 17–29.
- Yildirim A, Mavi A, Oktay M, Kara AA, Algur OF, Bilaloglu V (2000). Comparison of antioxidant and antimicrobial activities of *Tilia argentea* Desf Ex DC), Sage (*Salvia triloba* L.), and Black Tea (*Camellia sinensis*) extracts. *Journal of Agricultural and Food Chemistry*, 48: 5030-5034.

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